



Immunohistochemistry Protocol

Web: www.anbobio.com
www.dijibio.com

Order: order@dijibio.com
Support: support@dijibio.com

Tel: +86 519 8805 0026

Deparaffinization

1. Incubate slide at 60°C for 60 minutes.
2. Deparaffinize in Xylene for 10 minutes and repeat one more times.
3. Hydrate in 100% alcohol for 5 minutes, in 95% alcohol for 5 minutes, in 85% alcohol for 5 minutes, in 75% alcohol for 5 minutes.
4. Dip into Distill Water for 5 minutes.
5. Dip into PBS (pH 7.6), leave for 5 minutes, and repeat two times.

Antigen Retrieval

6. Put the slide into staining rack and lower into a container with 10 mM sodium citrate buffer solution (pH6.0), ensuring the slide is well immersed in sodium citrate buffer solution
7. Put the container into the microwave oven. Then high heat 5mins and medium heat 10-15mins
8. Cool the slide naturally to room temperature.
9. Dip into distilled water, leave for 5 minutes, and repeat two times.
10. Dip the slide in PBS for 5 minutes and repeat two times.
11. Immerse slides in 3% H₂O₂ (in fresh methanol) for 15 minutes at room temperature.
12. Wash with distilled water two times, 5 minutes each time.
13. Wash with PBS (pH 7.6) two times, 5 minutes each time.

Staining with Primary Antibody

14. Dilute primary antibody with 3% BSA in PBS. Cover the tissue section on the slide with diluted primary antibody (use 50 – 150µl for each slide).
15. Incubate at 37°C for 30 minutes or at room temperature for 60 minutes (The optimal incubation time, incubation temperature, and antibody dilution should be determined by the individual laboratory).
16. Wash with PBS two times, 5 minutes each time.

Staining with Secondary Antibody

17. Incubate with 100-200µl Polymer Enhancer. Incubate 30 minutes at 37°C.
18. Wash with PBS for 3 times, 5 minutes each time.
19. Incubate with 100-200µl Polymerized HRP and incubate 30 minutes at 37°C
20. Wash with PBS for 3 times, 5 minutes each time.
21. Add DAB solution and incubate 3-10 minutes (The reaction progress and the optimal time should be determine according to microscope).
22. Wash with distilled water for 2 times, 5 minutes each time.
23. Counterstain sections in hematoxylin if required, wash with distilled water. Immerse slides in 0.1% HCl-ethanol for 1-10 seconds, wash with distilled water.
24. Dehydrate through 95% ethanol for 1 minute, 100% ethanol for 2x3min, Xylene for 2x3min, and coverslip with mounting medium.